

Bacteriophage $\Phi 6$ as Surrogate and Human-Harmless Viruses to Study Anti-SARS-CoV-2 Approaches

Joana Barros^{1,2}, Maria Pia Ferraz^{1,2,3*}, Fernando Jorge Monteiro^{1,2,4}

¹IS – Instituto de Investigação e Inovação em Saúde, Universidade do Porto, Porto, Portugal

²INEB – Instituto de Engenharia Biomédica, Universidade do Porto, Porto, Portugal

³FCS/UFP – Faculdade de Ciências da Saúde, Universidade Fernando Pessoa, Porto, Portugal

⁴FEUP – Faculdade de Engenharia, Universidade do Porto, Porto, Portugal

Received: January 8, 2021; **Accepted:** January 20, 2021; **Published:** January 23, 2021

Citation: Barros J, Pia Ferraz M, Monteiro FJ (2021) Bacteriophage Phi 6 as Surrogate and Human-Harmless Viruses to Study Anti-SARS-CoV-2 Approaches. *Clar J Infect Dis Ther* 02 (02): 126–128.

Abstract

Given safety challenges in conducting laboratory work with highly infectious human coronaviruses (pathogenicity, genetic mutations rate, biosafety level 3 and 4 requirements), many researchers have valued the potential of bacteriophages as appropriate viral surrogate to measure humans enveloped virus' survival, transfer and removal. The use of phage $\Phi 6$ seems to be useful as coronavirus surrogate to assess the effectiveness of anti-SARS-CoV-2 approaches, providing important insights concerning COVID-19 pandemic and human public health.

Keywords: SARS-CoV-2; COVID-19; bacteriophage $\Phi 6$

Review

The 2019 Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) has emerged as a new respiratory pathogen and is responsible for large-scale morbidities and mortalities around the globe [1]. It is caused by a single positive-stranded RNA virus from the coronavirus (CoV) family of Coronaviridae, composed of four genera out of which α - and β -CoV can infect mammals including humans. SARS-CoV-2 is identified as β -CoV and is responsible for coronavirus disease 2019 (COVID-19) [1,2]. These viruses are wrapped in host cells derived lipid membranes where viral surface proteins are embedded. One of these surface proteins known as spike [S] protein protrudes out of membranes and gives a characteristic crown/halo-like appearance to the virus when observed under electron microscope, hence named coronavirus [1]. Once the virus gains entry into the respiratory tract, SARS-CoV-2 causes damage to

epithelial cells of the airways making lungs unable to clear dirt and mucus which can lead to pneumonia [1,2]. In extreme cases, patients experience a dramatic increase in the levels of pro-inflammatory chemokines and cytokines including IL-6 and TNF- α , a condition known as “cytokine storm”. This leads to the development of Acute Respiratory Distress Syndrome (ARDS), septic shock, metabolic acidosis, coagulation dysfunction, and even death [1,2].

Given safety challenges in conducting laboratory work with highly infectious human coronaviruses (pathogenicity, genetic mutations rate, biosafety level 3/4 (BSL-3 and BSL-4)), many researchers have valued the potential of bacteriophages (phages) as an appropriate viral surrogate to measure humans enveloped virus' survival, transfer and removal [3,4]. Phages seem to be good alternatives once they are relatively easy to produce in large quantities, and several purification procedures are laboratory available [3,5–8]. Bacterial viruses of biosafety

level 1 (BSL-1), pose no risk to humans, being safe for laboratory workers, and their study does not require specialized biocontainment precautions. Moreover, their similarity with eukaryote viruses allow cross-study comparisons, making them interesting models for aerovirology research [6,7,9,10]. In 2020, several studies have shown the potential of phage $\Phi 6$, non-pathogenic viruses that infect specifically bacterium *Pseudomonas syringae*, as a surrogate virus to study infections caused by enveloped human viruses [6,7,9,10]. For instance, Turgeon *et al* compared the effects of the aerosolization and sampling on the infectivity of 5 phages and 2 pathogenic viruses: MS2 (a single-stranded RNA [ssRNA] phage of the *Leviviridae* family), $\Phi 6$ (a segmented double-stranded RNA [dsRNA] phage of the *Cystoviridae* family), $\Phi X174$ (a single-stranded DNA [ssDNA] phage of the *Microviridae* family), PM2 (a double-stranded DNA [dsDNA] phage of the *Corticoviridae* family), PR772 (a dsDNA phage of the *Tectiviridae* family), human influenza A virus H1N1 (an ssRNA virus of the *Orthomyxoviridae* family), and the poultry virus Newcastle disease virus (NDV; an ssRNA virus of the *Paramyxoviridae* family)[11]. These authors showed that the behaviour of the influenza virus resembled that of phages PR772 and $\Phi 6$, providing critical information for the selection of appropriate phages models to mimic the behaviour of specific human and animal viruses in aerosols [11].

Phage $\Phi 6$ is a segmented RNA virus involved by a phospholipid envelope (fatty) with spike proteins at its surface, with ~80–100 nm size, structural features similar to several human viruses, namely Influenza (belongs to *Orthomyxoviridae* family), SARS-CoV-1, SARS-CoV-2 and Middle East Respiratory Syndrome-associated coronavirus (MERS-CoV) (belong to *Coronaviridae* family) [3,5–9]. Thus, phage $\Phi 6$ has been used as surrogate virus model to understand the relationship between environmental conditions and virus infectivity in order to improve strategies for predicting and controlling disease transmission, namely COVID-19 [3,5–9]. For instance, Casanova *et al* showed that recovery of phage $\Phi 6$ and *Influenza* virus from hands were comparable, with approx. 2–3 log₁₀ loss after using protein and non-ionic detergent-based eluent solutions [3]. These authors concluded that viruses' inactivation was probably due those solutions with capability to destabilize the fatty envelope structure,

a primary target for virus inactivation [3]. Dubuis *et al* showed that ozone at low concentration combined with high relative humidity was able to kill airborne viruses, such as phage $\Phi 6$ and murine norovirus MNV-1[12]. Rockey *et al* used phage $\Phi 6$ as surrogate model to evaluate the effectivity of heat and humidity treatments for N95 respirator de-contamination [9]. Buhr *et al* proved that phage $\Phi 6$ could be a useful indicator model to evaluate the inactivation and survival of an enveloped RNA virus on contaminated aircraft materials after exposure to hot, humid air [13]. Fedorenko *et al* showed that phage $\Phi 6$ presented a high survival rate in dry saliva deposited on glass surfaces, even when submitted to a wide range of relative humidity levels [6]. Phage $\Phi 6$ was considered a good model for virus respiratory pathogens, including SARS-CoV-2 [6].

Conclusion

Overall, the use of phage $\Phi 6$ may be useful as coronavirus surrogate to assess the effectiveness of anti-SARS-CoV-2 approaches, providing important insights concerning COVID-19 pandemic and human public health.

References

1. Zheng Y, Zhuang MW, Han L, Zhang J, Nan ML, Zhan P, *et al.* (2020) Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) membrane (M) protein inhibits type I and III interferon production by targeting RIG-I/MDA-5 signaling. *Signal Transduct Target Ther* 5: 299. [View]
2. Costela-Ruiz VJ, Illescas-Montes R, Puerta-Puerta JM, Ruiz C, Melguizo-Rodriguez L (2020) SARS-CoV-2 infection: The role of cytokines in COVID-19 disease. *Cytokine Growth Factor Rev* 54: 62–75. [View]
3. Casanova LM, Weaver SR (201) Evaluation of eluents for the recovery of an enveloped virus from hands by whole-hand sampling. *J Appl Microbiol* 118: 1210–6. [View]
4. Whitworth C, Mu Y, Houston H, Martinez-Smith M, Noble-Wang J, Coulliette-Salmond A, *et al.* (2020) Persistence of Bacteriophage Phi 6 on Porous and Nonporous Surfaces and the Potential for Its Use as an Ebola Virus or Coronavirus Surrogate. *Appl Environ Microbiol* 86: 01482–20. [View]
5. Prussin AJ 2nd, Schwake DO, Lin K, Gallagher DL, Buttlng L, Marr LC (2018) Survival of the Enveloped

- Virus Phi6 in Droplets as a Function of Relative Humidity, Absolute Humidity, and Temperature. *Appl Environ Microbiol* 84: 00551–18. [[View](#)]
6. Fedorenko A, Grinberg M, Orevi T, Kashtan N (2020) Survival of the enveloped bacteriophage Phi6 (a surrogate for SARS-CoV-2) in evaporated saliva microdroplets deposited on glass surfaces. *Sci Rep* 10: 22419. [[View](#)]
 7. Fedorenko A, Grinberg M, Orevi T, Kashtan N (2020) Virus survival in evaporated saliva microdroplets deposited on inanimate surfaces. *bioRxiv* :2020.06.15.152983. [[View](#)]
 8. Vatter P, Hoenes K, Hessling M (2020) Photoinactivation of the Coronavirus Surrogate phi6 by Visible Light. *Photochem Photobiol* 2020. [[View](#)]
 9. Rockey N, Arts PJ, Li L, Harrison KR, Langenfeld K, Fitzsimmons WJ, *et al.* (2020) Humidity and Deposition Solution Play a Critical Role in Virus Inactivation by Heat Treatment of N95 Respirators. *mSphere* 15: 00588–20. [[View](#)]
 10. Gendron L, Verreault D, Veillette M, Moineau S, Duchaine C (2010) Evaluation of Filters for the Sampling and Quantification of RNA Phage Aerosols. *Aerosol Science and Technology* 44: 893–901. [[View](#)]
 11. Turgeon N, Toulouse MJ, Martel B, Moineau S, Duchaine C (2014) Comparison of Five Bacteriophages as Models for Viral Aerosol Studies. *Appl Environ Microb* 80: 4242–50. [[View](#)]
 12. Dubuis ME, Dumont-Leblond N, Laliberte C, Veillette M, Turgeon N, Jean J, *et al.* (2020) Ozone efficacy for the control of airborne viruses: Bacteriophage and norovirus models. *Plos One* 15. [[View](#)]
 13. Buhr TL, Young AA, Borgers-Klonkowski E, Kennihan NL, Barnette HK, Minter ZA, *et al.* (2020) Hot, Humid Air Decontamination of Aircraft Confirmed That High Temperature and High Humidity Are Critical for Inactivation of Infectious, Enveloped Ribonucleic Acid (RNA) Virus. *Front Bioeng Biotech* 8. [[View](#)]

***Corresponding author:** Maria Pia Ferraz, Faculdade de Ciências da Saúde, Universidade Fernando Pessoa, Rua Carlos da Maia, 296, 4200-150 Porto, Portugal;

Email: mpferraz@ufp.edu.pt